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Phytoconstituents and *In Vitro* Free Radical Scavenging Potential of *n*-Hexane and Aqueous Fractions of *Cucurbita maxima* and *Leptadenia hastata*

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Abstract: The present study explored the phytoconstituents and radical scavenging activity of the respective *n*-hexane and aqueous fractions of *Cucurbita maxima* (CMHF and CMAF) and *Leptadenia hastata* (LHHF and LHAF) for potential application in oxidative stress-related ailments. The phytoconstituents were qualitatively determined and characterized using Fourier-transform Infrared (FTIR), while the antioxidant activity was determined *in vitro*. Alkaloids were present in only the aqueous fractions of *C. maxima* and *L. hastata*, while saponins, steroids, and flavonoids were detected in all the fractions. The FTIR revealed the presence of functional groups, including alcohols, sulfonates, alkenes, alkanes, amines, and aromatics in both plant fractions. The LHHF (35.53 ±2.11 ascorbic acid (AA) equivalent µg/mL) exhibited a significantly (p<0.05) higher total reducing power (TRP) than all the other fractions. The CMHF (69.11 ±2.56 AAE µg/mL) demonstrated a significantly (p<0.05) higher total antioxidant capacity (TAC) than all the other fractions. For the ferric thiocyanate (FTC) assay, the highest inhibition was exhibited by LHHF (79.78 ± 3.24%), significantly (p<0.05) higher than AA (26.46 ± 2.12%), CMHF (69.77 ± 3.16%), and CMAF (43.80 ± 2.12%). In the thiobarbituric acid assay, the lowest MDA concentration was exhibited by the CMHF (0.07 ±0.01 nmol/mL), significantly (p<0.05) lower than all the other fractions and ascorbic acid. Conclusively, the *n*-hexane fraction of both plants presents potential sources of novel antioxidant compounds with significant free radical scavenging and anti-lipid peroxidation activities, applicable in ailments linked to oxidative stress.

Introduction

Oxidative stress (OS) defines a redox imbalance in a cell that is linked to several ailments, including cancer and diabetes. A state of redox imbalance is established in a cell when the number of free radicals released exceeds the capacity of the inherent antioxidant system of the body, notably in cancer and diabetes (1, 2). This is further exacerbated by the progression into an inflammatory statepromoted by the inflammatory cytokines (2, 3). Free radicals, including the reactive oxygen species (ROS) and reactive nitrogen species (RNS), are normal components of metabolic pathways and processes in a cell, such as phagocytosis, mitochondrial respiratory chain, and arachidonic acid metabolism (4). However, the antioxidant systems within the cell, such as glutathione peroxidase, catalase, and superoxide dismutase, neutralize and prevent their damaging effects on cellular components, including the DNA and membrane lipids (5). There are various free radicals, including peroxyl (ROO·), hydroxyl (·OH), alkoxy (-OR), nitric oxide (•NO), and superoxide ($O_2^{\bullet -}$). Additionally, hydrogen peroxide (H_2O_2), although not a radical, is often involved in oxidative processes (6). Once released without subsequent neutralization, these free radicals cause

lipid peroxidation, protein modification, DNA fragmentation and damage, and cell death (1, 2). Thus, the need for free radical neutralization highlights the significant roles played by both endogenous and exogenous antioxidants in health and diseases.

Antioxidants, including the exogenous and their endogenous counterparts, are vital components of the cell, and their role is coupled to many pathways and cellular processes. Antioxidants act by donating or accepting electrons to free radicals, thereby neutralizing and preventing their associated effects (7). Exogenous sources of antioxidants entail compounds including phytochemicals with electron-donating capabilities, such as flavonoids, made up of the hydroxyl, carbonyl (oxo), and catechol groups within their structure. Moreover, the conjugated double bond within the flavonoid structure contributes to the antioxidant capabilities of these compounds (8, 9). Alkaloids, including allocryptopine, berberine, caffeine, and chelerythrine, exert their antioxidant properties by modulating pathways linked to OS (10). Allocryptopine modulates Akt/glycogen synthase kinase-3β (GSK-3β) signaling, significantly reducing OS-induced tau protein hyperphosphorylation. At the same time, berberine decreases endoplasmic reticulum (ER) stress and OS by suppressing ER PKR-like kinase (PERK)/eukaryotic translation initiation factor 2α (eIF2α) signalling-mediated beta-secretase 1 (BACE-1) translation (11, 12). Saponins, another class of phytochemicals, also exert their radical scavenging effects by modulating signaling cascade genes linked to OS (13). In a previous report, saponins were observed to upregulate antioxidant-related genes in human embryonic kidney 293 cells by activating nuclear factor erythroid 2-related factor 2 (13). Although these phytochemicals are attributed with antioxidant activities in cells, their radical scavenging potential presents a gap to explore in plant-based drugs for therapeutic application in OS-linked ailments.

Medicinal plants present a vast reserve of phytochemicals with different pharmacological activities, working individually or synergistically, including flavonoids, alkaloids, and saponins (14-18). In folkloric medicine, plant-based drugs prepared in different formulations are utilized against different ailments to achieve a desired therapeutic effect. *L. hastata* has been employed in traditional medicine to treat hypertension, erectile dysfunction, trypanosomiasis, diabetes mellitus, rhinitis (catarrh), skin diseases, and acute rhinopharyngitis (19, 20). *C. maxima* treats constipation, anemia, intestinal parasites, wounds, renal ailments, cataracts, and blood pressure (21, 22). The antioxidant activity of different plants, attributed to their phytoconstituents, was previously investigated via *in vitro* and *in silico* techniques (15, 16, 23-26). Some of these phytoconstituents were reported to be present in *C. maxima* and *L. hastata*, respectively. However, the extraction of these phytochemicals depends on the solvent employed during extraction. Moreover, the fractionation allows the partitioning of the phytochemicals based on solvent polarity. Previous studies focused on the *in vitro* antioxidant activities and characterization of the crude extracts of these plants via the gas-chromatography mass spectroscopy technique and 2,2-diphenyl-1-picrylhydrazyl assays, respectively (27, 28). However, in the present study, we characterized the phytochemicals and determined the antioxidant activity of the *n*-hexane and aqueous fractions of *C. maxima* and *L. hastata* seed and leaf, respectively, for potential application in oxidative stress-related ailments.

Methodology

Materials

C. maxima and *L. hastata* were collected from the bypass of Jimeta Area of Adamawa State, Nigeria. A Forest Technologist from the Forestry Technology Department of Adamawa State Polytechnic, Yola, identified the plant. A voucher specimen (No. ASP/FT/23/012) was deposited at the Department. The seed and leaf of *C. maxima* and *L. hastata* were air-dried and ground to powder using a blender to obtain the powdered sample.

Extraction and Fractionation

Exactly 1 kg of the samples were separately macerated in 3 L of 75% (v/v) aqueous ethanol for 7 days in an air-tight container with daily agitation. This was followed by filtration using Whatman no. 1 filter paper to obtain the filtrate, which was concentrated to dryness with a rotary evaporator (Buchi Rotavapor R-200) under reduced temperature and pressure to obtain the crude extracts (72 and 165 g of *C. maxima* and *L. hastata*, respectively). Exactly 50 g of *C. maxima* crude extract was completely dissolved in 200 mL of distilled water and transferred to a 500 mL separating funnel. Then, an equal volume of n-hexane and vigorous shaking was added for complete partitioning and collection of the *n*-hexane layer. The aqueous layer was continuously washed with an equal volume of *n*-hexane until a clear *n*-hexane layer was obtained. The *n*-hexane and aqueous layers were dried *via* the same procedure for the crude extracts to obtain 19 and 29 g of the *n*-hexane (CMHF) and aqueous (CMAF) fractions of *C. maxima*, respectively. For the *L. hastata*, 100 g of the crude extract was subjected to the same treatment to obtain 44 and 62 g of the *n*-hexane (LHHF) and aqueous (LHAF) fractions, respectively.

Phytochemical Detection

The method previously described by Evans (29) was used to detect the presence of alkaloids, saponins, steroids, glycosides, terpenoids, and flavonoids.

FTIR Characterization

The functional group identification was carried out using Perkin-Elmer FTIR (series 100), according to the method described previously (30). The IR spectrum Table was used to identify the functional groups by interpreting and comparing the peaks.

Radical Scavenging Potential

The radical scavenging potential of the extracts, including the total reducing power (TRP), total antioxidant capacity (TAC), ferric thiocyanate assay (FTC), and thiobarbituric acid assay (TBA) were determined as follows:

TRP

The TRP was determined based on the method described by Oyaizu (31). Briefly, 0.25 mL of 100 µg/mL of the sample was mixed with 0.625 mL of 0.2 M phosphate buffer (pH 6.6) and 0.625 mL of 1% (w/v) potassium ferrocyanide. The mixture was incubated for 50 min at 50°C, followed by adding 0.625 mL of 10% (w/v) trichloroacetic acid (TCA) and 10 min centrifugation at 3000 rpm. Precisely 1.8 mL of the upper layer was collected and mixed with an equal volume of distilled water. The absorbance was read at 700 nm against a blank (without sample) after adding 0.36 mL of 0.1% $FeCl_3$. Moreover, varying concentrations (20-100 µg/mL) of ascorbic acid (AA) mixed with the reagents were used to obtain the AA calibration curve. The result was expressed as AA equivalent (AAE) in µg/mL of the mean of triplicate determinations.

TAC

The TAC was determined following the protocol Prieto et al. described (32). Briefly, 0.5 mL of 300 µg/mL of the sample was mixed with phosphomolybdate reagent. The mixture was capped and incubated for 10 min at 95°C. The absorbance of the sample was read at 695 nm after cooling against a blank (without the sample). Furthermore, varying concentrations (20-100 µg/mL) of AA mixed with the reagent were used to obtain the AA calibration curve. The result was expressed as AAE in µg/mL of the mean of triplicate determinations.

FTC

The FTC assay was used to determine the effect of the extract on the early stage of lipid peroxidation according to the protocol of Kikuzaki and Nakatani (33). Briefly, 4 mL of 1 mg/mL of the sample dissolved in absolute ethanol was mixed with 0.05 M pH 7 phosphate buffer and 3.9 mL distilled water, capped, and incubated in a dark oven at 40°C for 10 min. Exactly 0.1 mL of the solution was mixed with 0.1 mL of 30% (w/v) ammonium thiocyanate, 9.7 mL of 75% (v/v) ethanol, and 0.1 mL of 0.02 M ferrous chloride in 3.5% (v/v) HCl. A blank solution with only the reagents was used as a control, while AA was standard. The absorbance was read immediately and after every 24 h at 532 nm until the maximum absorbance of the control was observed. The percentage inhibition of lipid peroxidation was evaluated from Equation 1. The result was expressed as the mean of triplicate determination.

$$\%Inhibition = 100 - \left(\frac{At}{Ac} \times 100\right) \quad \textbf{Equation 1}$$

Where At = absorbance of sample, while Ac = absorbance of control.

TBA

The effect of the extracts on the late stage of lipid peroxidation (MDA formation) was determined as described by Kikuzaki and Nakatani (33). Briefly, 1 mL of the prepared sample in the FTC method was mixed with 2 mL each of 20% (w/v) TCA and 0.5% (w/v) TBA. The mixture was capped and incubated in a water bath for 20 min at 90°C, followed by centrifugation at 3000 rpm after cooling. The absorbance of the supernatant was read at 532 nm on the last of the FTC method. The MDA concentration was evaluated from Equation 2.

$$MDA\,concentration = \left(\frac{OD}{EC}\, x\, V_s\right) \quad \textbf{Equation 2}$$

Where OD = absorbance of the sample, while EC = extinction coefficient.

Statistics

The results were evaluated using the Statistical Package for Social Sciences (SPSS) version 22 software and expressed as the mean of ± standard error of the mean of triplicate determinations (± SEM). The group difference was evaluated by One-way analysis of variance, followed by Tukey's multiple comparison test.

Results

Phytochemical Detection

The phytoconstituents detected in the CMHF, LHHF, CMAF, and LHAF are shown in **Table 1**. Alkaloids were present in only the aqueous fractions of *C. maxima* and *L. hastata,* while saponins, steroids, and flavonoids were detected in all the fractions. Moreover, glycosides and terpenoids were absent in all the fractions.

Table 1. Phytochemicals detected in the *n-hexane* and aqueous fractions of *C. maxima* and *L. hastata.*

Phytochemicals	CMHF	LHHF	CMAF	LHAF
Alkaloids	-	-	+	+
Saponins	+	+	+	+
Steroids	+	+	+	+
Glycosides	-	-	-	-
Terpenoids	-	-	-	-
Flavonoids	+	+	+	+

Note: (+) = present and (-) = absent

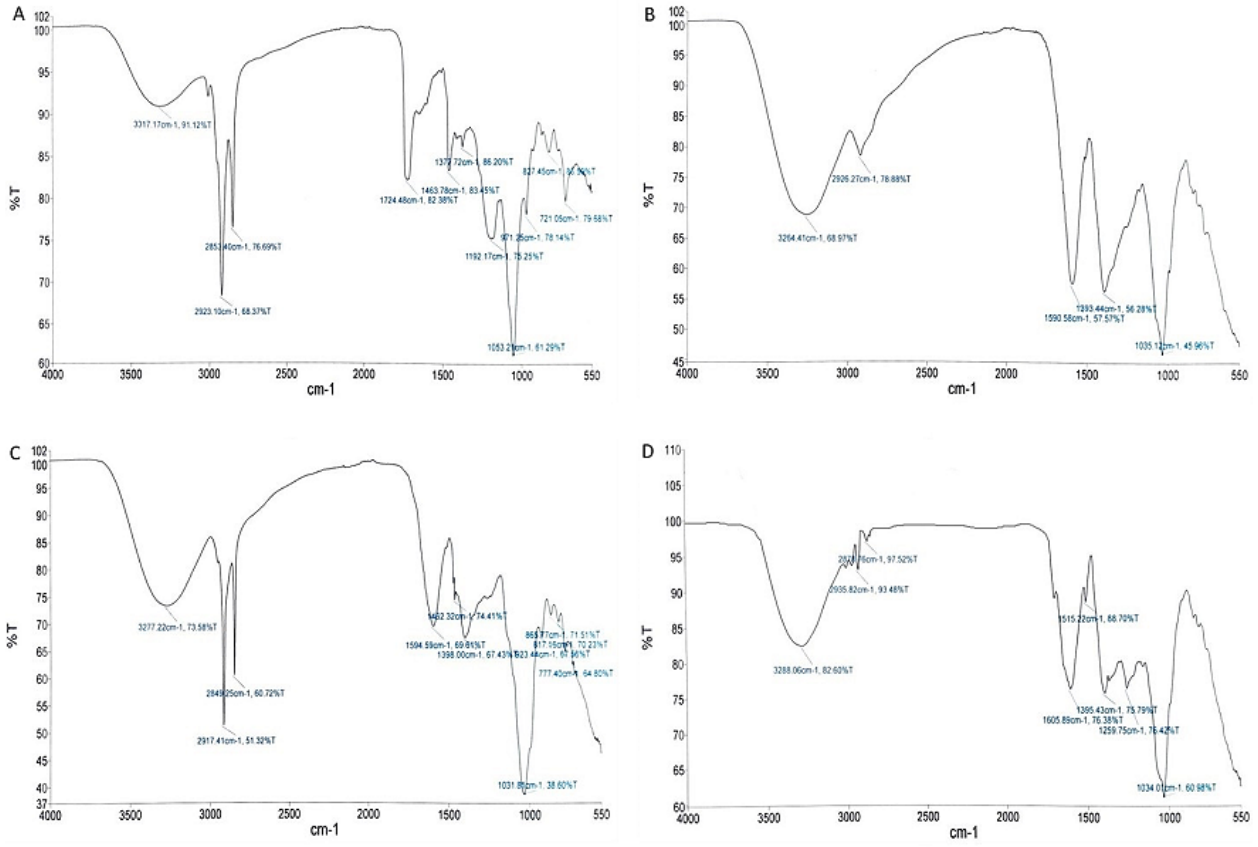

Figure 1. FTIR spectrum of *n*-hexane and aqueous fractions of *C. maxima* and *L. hastata*; (A) CMHF, (B) LHHF, (C) CMAF, and (D) LHAF.

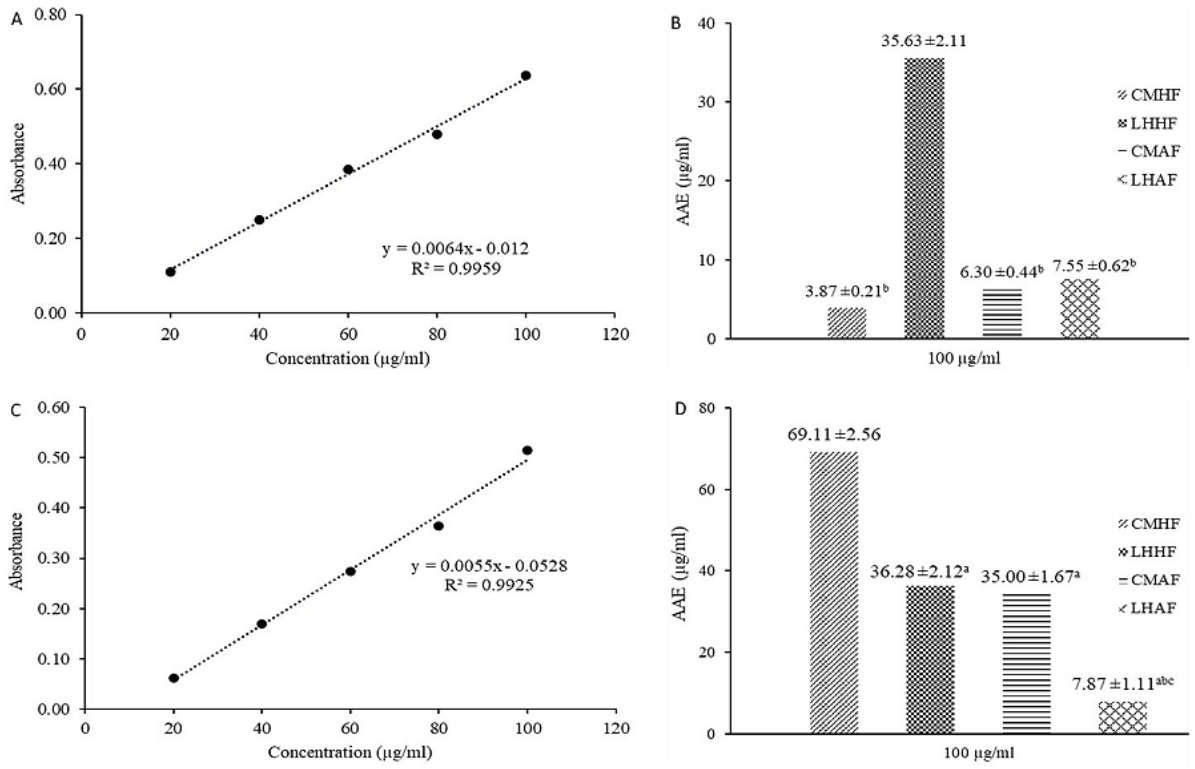

Figure 2. TRP and TAC of *C. maxima* and *L. hastata*; (A) AA TRP calibration curve, (B) TRP, (C) AA TAC calibration curve, and (D) TAC. Values with a, b, and c superscripts are significantly ($p<0.05$) lower than CMHF, LHHF, and CMAF, respectively.

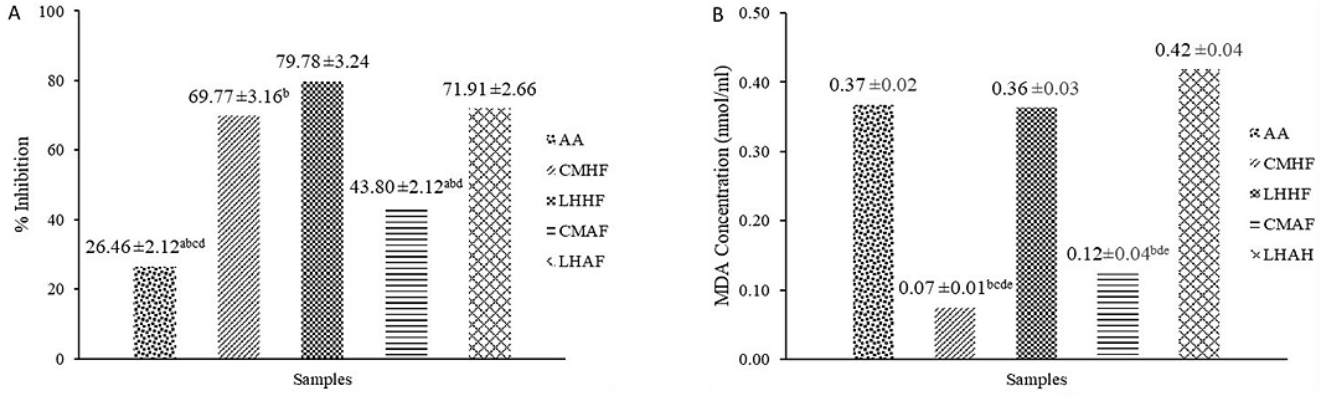

Figure 3. Anti-lipid peroxidation potential of *C. maxima* and *L. hastata*; (A) FTC assay, (B) TBA assay. Values with a, b, c, d, and e superscripts are significantly (p<0.05) lower than CMHF, LHHF, CMAF, LHAF, and AA, respectively.

FTIR Characterization

The FTIR spectrum of the CMHF, LHHF, CMAH, and LHAF is presented in **Figure 1**, showing the absorption peaks. A total of 11 peaks were detected, including four in the group frequency region. For the CMHF, the first absorption peak was at 3317.17 cm^{-1}, corresponding to the O-H stretching frequency of the alcohol compounds. The second (2923.10 cm^{-1}) and third (2853.40 cm^{-1}) peaks at the frequency regions correspond to the N-H stretching frequency of amine salt, while 1724.48 cm^{-1} corresponds to the C=O stretching of the α, β-unsaturated esters. In the fingerprint region, the peak at 1463.78 (C-H bending), 1377.72 (S=O stretching), 1192 (C-O stretching), 971.25 (C=H bend), 818.53 (C-H bending), and 721.05 cm^{-1} fingerprint regions corresponds to the alkane, sulfonates, ester, primary alcohol, alkene, disubstituted alkanes, aromatics, respectively. A total of 5 peaks were detected for the LHHF, including two in the group frequency region. The first absorption peak was at 3264.41 cm^{-1}, corresponding to the O-H stretching frequency of the alcohol compounds. The second (2926.27 cm^{-1}) peak at the frequency regions corresponds to the N-H stretching frequency of amine salt. In the fingerprint region, the peak at 1590.58 (N-H bending), 1398.44 (S=O stretching), and 1027.37 (C-N stretch) fingerprint regions correspond to sulfonates and amine groups, respectively.

A total of 11 peaks were detected for the CMAH, including three in the group frequency region. The first absorption peak was at 3277.22 cm^{-1}, which corresponded to the O-H stretching frequency of the alcohol compounds. The second (2917.41 cm^{-1}) and third (2849.25 cm^{-1}) peaks at the frequency regions correspond to the N-H stretching frequency of amine salt. In the fingerprint region, 1594.59 (N-H bending), 1462.32 (C-H bending), 1398.00 (S=O stretching), 923.44 (C-H bend), 817.16 (C=C bending), and 777.40 cm^{-1} (C-H bend) fingerprint regions corresponds to the amine, alkane, sulfate, monosubstituted alkenes, trisubstituted alkenes, and ortho aromatics, respectively. A total of 8 peaks were detected for the LHAF, including four in the group frequency region. The first absorption peak was at 3288.06 cm^{-1}, corresponding to the O-H stretching frequency of the alcohol compounds. The second (2935.82 cm^{-1}) and third (2878.76 cm^{-1}) peaks at the frequency regions correspond to the N-H stretching frequency of amine salt. In the fingerprint region, the peak at 1605.89 cm^{-1} corresponds to the C=C frequency of conjugated alkenes. The 1515.22 (N-O stretching), 1395.43 (S=O stretching), 1259.75 (C-N stretching), and 1027.37 (C-N stretch) fingerprint regions correspond to the nitrogen compound, sulfate, alkyl aryl ether, and amines, respectively.

Radical Scavenging Potential

TRP

The AA calibration curve and TRP of the CMHF, LHHF, CMAF, and LHAF are shown in **Figure 2**, depicting their AA equivalent total reducing power. The LHHF (35.53 ± 2.11 AAE µg/mL) exhibited a significantly (p<0.05) higher TRP than all the other fractions. Additionally, there was no significant (p>0.05) difference between the TRP of CMHF (3.87 ± 0.21 AAE µg/mL), CMAF (6.30 ± 0.44 AAE µg/mL), and LHAF (7.55 ± 0.62 AAE µg/mL) though the latter was slightly higher.

TAC

Figure 2 shows the AA calibration curve and TAC of the CMHF, LHHF, CMAF, and LHAF, revealing their AA equivalent total antioxidant capacity. The CMHF (69.11 ± 2.56 AAE µg/mL) demonstrated a significantly (p< 0.05) higher TAC than all the other fractions. Moreover, the LHAF (7.87 ± 1.11 AAE µg/mL) was significantly (p< 0.05) lower than the LHHF (36.28 ± 2.12 AAE µg/mL) and CMAF (35.00 ± 1.67 AAE µg/mL).

FTC

The anti-lipid peroxidation potential of CMHF, LHHF, CMAF, and LHAF is presented in **Figure 3**, showing the percentage inhibition. The highest inhibition was exhibited by the LHHF (79.78 ± 3.24%), significantly

(p<0.05) higher than AA (26.46 ± 2.12%), CMHF (69.77 ± 3.16%), and CMAF (43.80 ± 2.12%). The CMHF (69.77 ± 3.16%) exhibited a significantly (p<0.05) higher inhibition than AA (26.46 ± 2.12%) and CMAF (43.80 ± 2.12%). Moreover, the LHAF (71.91 ± 2.66%) was significantly (p<0.05) higher than AA (26.46 ± 2.12%) and CMAF (43.80 ± 2.12%).

TBA

Figure 3 shows the anti-lipid peroxidation potential of CMHF, LHHF, CMAF, and LHAF, depicting the MDA concentration. The lowest MDA concentration was exhibited by the CMHF (0.07 ± 0.01 nmol/mL), significantly (p<0.05) lower than AA (0.37 ± 0.02 nmol/mL) and all the fractions. Moreover, the CMAH had a significantly (p<0.05) lower MDA concentration than AA (0.37 ± 0.02 nmol/mL), LHHF (0.36 ± 0.03 nmol/mL), and LHAF (0.42 ± 0.04 nmol/mL).

Discussion

Phytochemicals confer medicinal properties in plants and are implicated in the therapeutic effects of such plants. Different plants contain variable phytoconstituents, thus exhibiting different pharmacological effects. In our study, fractionation of the crude extract separated the phytochemicals based on polarity via the partitioning technique, which detected alkaloids in only the aqueous fractions. The polarity of solvents has been reported to influence the class of phytochemicals extracted, which also influences the pharmacological activities of the extracts, including their antioxidant potential (34). The phytochemicals detected in our study were previously implicated with antioxidant properties (10, 35, 36).

Moreover, the presence of these phytochemicals was previously reported (37). Mwangi et al. (37) reported similar results to the present study, detecting alkaloids, saponins, steroids, and flavonoids without terpenoids in *C. maxima*. In another study, Pal et al. (38) reported similar results to our study, detecting alkaloids and saponins in the aqueous seed extract of *C. maxima*. In their study, Omeh et al. (39) detected the presence of alkaloids, saponins, and flavonoids in *L. hastata* in addition to terpenoids and glycosides, slightly disagreeing with our report. However, the report of Maria and Olorukooba (40) on alkaloids, saponins, steroids, and flavonoids in *L. hastata* is concurrent with our result.

The radical scavenging capabilities of plant-based drugs are attributed to the presence of compounds and functional groups with electron-donating potential, neutralizing the free radicals generated from different pathways in cells. Additionally, these compounds act as ligands, modulating cell pathways and processes (11-13). Thus, detecting these functional groups in plants might offer antioxidant properties, including free radical scavenging effects. In our report, some of the functional groups detected include alkane, sulfonates, esters, alcohols, alkenes, disubstituted alkanes, aromatics, carboxylic acids, and amine salts which might be responsible for the radical scavenging of the plants. Chen et al. (41) reported the antioxidant activity of carboxylic acid and alcohol groups *via* a structure-activity relationship study, depicting enhanced antioxidant activity attributed to these groups. In another study, Fikriya and Cahyana (42) reported enhanced antioxidant activity of quinoline-4-carboxylic acid, synthesized through the Pfitzinger reaction *via* the reaction of a ketone with isatin. The result showed an increased antioxidant activity compared to the parent compound isatin, attributed to carboxylic acid and aromatic ring presence. Li et al. (43) reported a similar result with the presence of the carboxylic acid group in 1,8-Dihydroxyanthraquinone, leading to enhanced free radical scavenging compared to its analogs upon photoexcitation. The functional groups reported in our study were previously reported in other studies. Andronie et al. (44) reported the presence of alcohol, amine, alkane, and alkene groups in *C. maxima* seeds *via* the FTIR technique. Mwangi et al. (37) reported similar findings, identifying alcohol, amine, alkane, and alkene groups in *C. maxima* seeds. In another study, alcohol, amine, alkane, and alkene groups were detected in *L. hastata* leaf concurrently with our report (45).

The antioxidant activity is partly capable of scavenging free radicals by neutralizing and mitigating their effect on cellular components, including lipids and DNA (7, 46). This might serve as a first defensive strategy against free radicals in ailments associated with increased free radical generation, including diabetes, cancer, and neurodegenerative diseases. Thus, plant-based therapy to scavenge and neutralize the free radicals might serve beneficial effects. The ferric-reducing antioxidant capacity method is principled on reducing ferric to ferrous ions in the presence of the sample, yielding a directly proportional intense-blue color. Thus, the higher the absorbance, the higher the antioxidant capacity. A similar principle is followed for the total antioxidant capacity where Mo (VI) is reduced to Mo (V) by the antioxidant compounds present in the sample, which then forms a green phosphate/Mo(V) complex with maximum absorption at 695 nm. The principle behind this method relies on the ability of antioxidant molecules to donate electrons or hydrogen atoms to reactive species, thereby preventing oxidative damage. In our study, the superior total reducing power exhibited by the LHHF might be attributed to the presence of electron-donating compounds absent in the other fractions.

Interestingly, the CMHF showed superior total antioxidant capacity *via* the phosphomolybdate method. This variation might be attributed to the

specificity of the antioxidant compounds present in the two fractions with preferences towards the ferric ions (Fe^{3+}) or Mo (VI), demonstrating effectiveness to either assay (47). Further, AA exhibited superior TRP and TAC in our result, which correlates with the result reported by Kushawaha et al. (48) and Monica et al. (49). The TAC for the CMAF reported in our study was lower than the value reported by Kar et al., (28) and Tijjani et al., (50) for the crude aqueous extract of *C. maxima* which might be attributed to the absence of some compounds partitioned in the CMHF.

The FTC and TBA methods explore the early and end stages of lipid peroxidation, respectively, with the latter focused on inhibiting malonaldehyde (MDA) formation. MDA is often regarded as a biomarker of oxidative stress and the end product of lipid peroxidation, as increased lipid peroxidation leads to increased MDA formation. Although the LHHF showed the highest lipid peroxidation inhibition in our study, the CMHF exhibited the least MDA concentration, thus demonstrating superior activity against MDA formation. Kushawaha et al. (48) reported a comparable anti-lipid peroxidation activity of the aqueous extract of *C. maxima* seed with AA, which partially agrees with the result of the aqueous fraction in our study.

Conclusion

In the present study, the main objective was to explore the phytoconstituents and the free radical scavenging effect of n-hexane and aqueous fractions of C. maxima and L. hastata. The *n*-hexane fractions of both plants exhibited significant antioxidant activity, with *L. hastata* and *C. maxima* demonstrating superior total reducing power and total antioxidant capacity, respectively. Moreover, a similar observation was made for the anti-lipid peroxidation of the plants with superior activity exhibited by the *n*-hexane fractions of *C. maxima* and *L. hastata via* the FTC and TBA assays, respectively. These antioxidant activities might be attributed to the phytoconstituents with antioxidant functional groups identified in the FTIR analysis. Conclusively, both plants present a potential source of novel antioxidant compounds, with the *n*-hexane fractions demonstrating superior free radical scavenging and anti-lipid peroxidation activities. Thus, these plants can be applicable in the phytotherapy of ailments linked to oxidative stress, including diabetes, cancer, and aging-related diseases.

Abbreviations

AA = Ascorbic acid; CMAF = *Cucurbita maxima* Aqueous Fraction; CMHF = *Cucurbita maxima n*-hexane Fraction; FTC = Ferric Thiocyanate; FTIR = Fourier-transform Infrared; LHHF = *Leptadenia hastata n*-hexane Fraction; LHAF = *Leptadenia hastata* Aqueous Fraction; MDA = Malonaldehyde; OS = Oxidative Stress; ROS = Reactive Oxygen Species; RNS = Reactive Nitrogen Species; TAC = Total Antioxidant Capacity; TBA = Thiobarbituric Acid Assay; TRP = Total Reducing Power.

Declarations

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Conflict of Interest
The authors declare no conflicting interest.

Data Availability
The unpublished data is available upon request to the corresponding author.

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Not applicable.

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